

Functional Analysis of the *Drosophila* H731 Protein by the Yeast Two-hybrid System

Abstract *Drosophila* H731 (dH731) is the homologue of mouse Pcd4, which is implicated in apoptosis and tumorigenesis. In order to investigate the possible cellular functions of the dH731 protein, I attempted to find out the interacting proteins of dH731 as well as to elucidate the functional relationship between dH731 and its interacting proteins. By yeast two-hybrid screening and database search, nine different dH731-interacting clones were isolated from 5×10^6 yeast clones. Five of them encode novel *Drosophila* proteins, and two of the five have a C2H2 zinc finger motif which is a putative DNA binding domain. The other four proteins are dH731, dUBC9, rpL23a and eIF4a. Therefore, the data suggest that dH731 protein can bind to itself, forming homodimers or multimers. The mRNA expression patterns of the dH731 gene and its interacting genes were examined by whole-mount in situ hybridization of the developing embryos. They were all localized in the central nervous system and partially overlapped in other tissues, including gonads. Taken together, it is conceivable that dUBC9 may help the dH731 protein enter the nucleus to interact with rpL23a and two other C2H2 zinc finger-containing proteins, and that the dH731 protein may play a role in regulation of transcription or cell cycle progression during the development of the nervous system and the reproductive organ.

1. Introduction The *Drosophila* H731 protein (dH731) was identified through its interaction with Numb. Numb plays an important role in determining cell fate during neural development (1)dH731 encodes 509 amino acid residues. The mouse Pcd4 protein, which may inhibit tumor formation (2)shows 41% identity to dH731, but the functions of dH731 have not been elucidated. Therefore, I am very interested in the functions of the dH731 protein during embryonic development. The yeast two-hybrid system (3) (Fig. 1) was used to search for the interacting proteins of dH731, and nine different proteins were found. The functional relationship between dH731 and its interacting proteins was discussed.

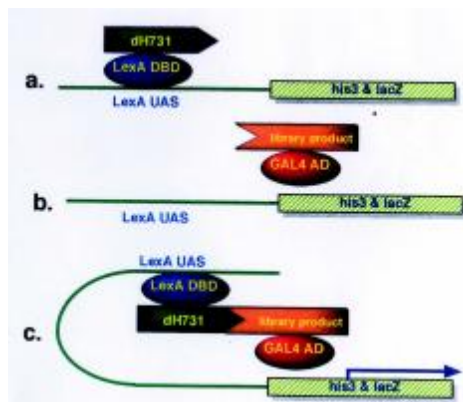


Fig. 1 Yeast two-hybrid system

2. Materials and Methods

1. DNA cloning The DNA sequences of dH731, the full length and the C-terminus, were cloned into pBHA which contains sequences encoding the LexA DNA binding protein. The dH731 sequence was cloned downstream of LexA sequence (Fig.2).

2. Yeast two-hybrid screening pBHA-dH731 and pGAD10-library were co-transformed into the L40 yeast with a heat shock pulse of 42°C for 15 min. Positive transformants were confirmed repeatedly for three times of growth (for histidine synthesis) and X-gal assays (for β -galactosidase activity). To prevent more than one plasmid in a single yeast colony, pGAD10-library plasmids were extracted from positive candidates, and transformed into the HB101 *E. coli*.

3. Grouping and retransformation Since these positive candidates might contain DNA sequence from the same gene, cross hybridization by dot blotting was used to group the candidates. Non-radioactive DIG-labeled probes were generated and hybridization was performed at 65°C . Representatives from each

group were retransformed into yeast L40 with pBHA-lamin to exclude non-specific interactors. Transformants that did not turn on the reporter genes with pBHA-lamin were the true positive.

4. Sequencing and database search The DNA sequences of dH731-interacting clones were subjected for sequencing. The resulting sequences were processed with softwares DNA Strider 1.2 and Gene Work 2.0, and were then compared to DNA sequence databases Berkeley Drosophila Genome Project, WWW BLAST at National Center for Biotechnology Information, Welcome to PubMed and ISREC ProfileScan Server.

5. Whole-mount embryo in situ hybridization The mRNA expression patterns of dH731 and its interacting genes were examined by whole-mount in situ hybridization of the developing embryos. The mRNA of these genes was hybridized by DIG-labeled probes which were recognized by anti-DIG-alkaline phosphatase (AP). These embryos were then subjected for color reactions to reveal the expression pattern.

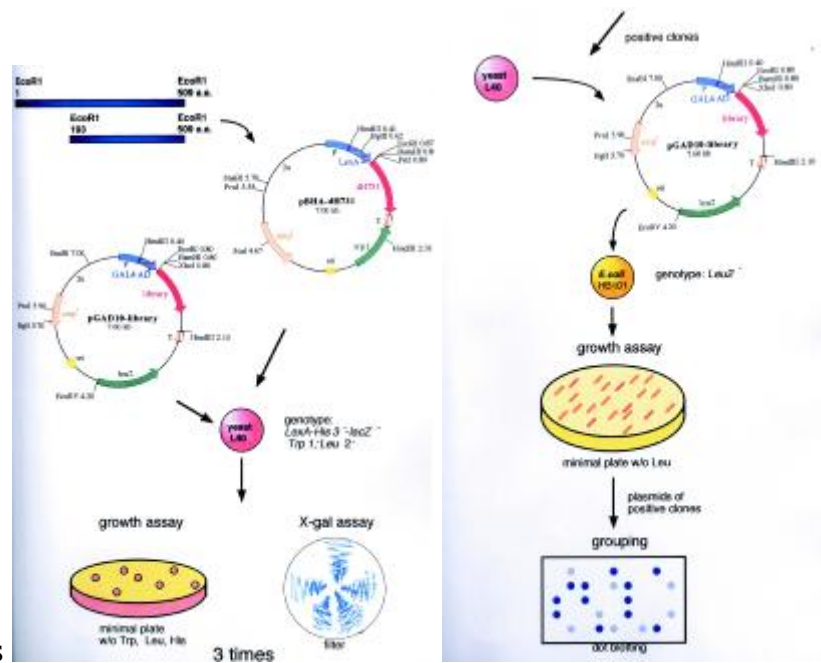
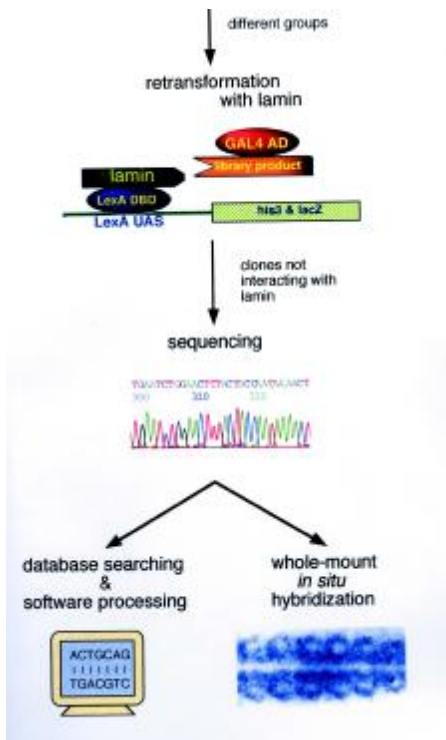


Fig. 2 Schematic procedures



3.Results 1. DNA cloning The DNA cloning results were examined by agarose gel electrophoresis (Fig. 3). The three DNA sequence fragments of the dH731 gene were cloned to be in frame with LexA protein. These DNA inserts were also confirmed by sequencing.

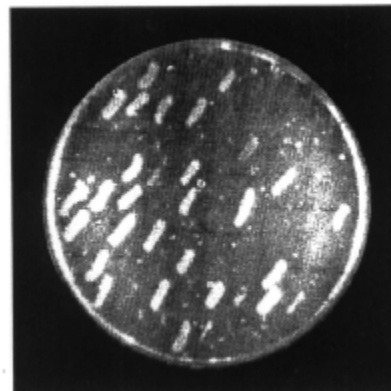
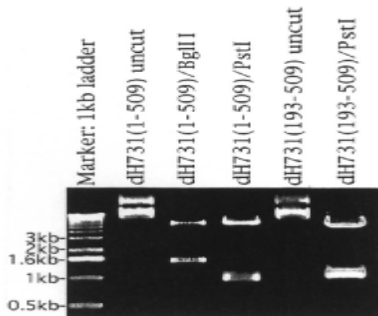
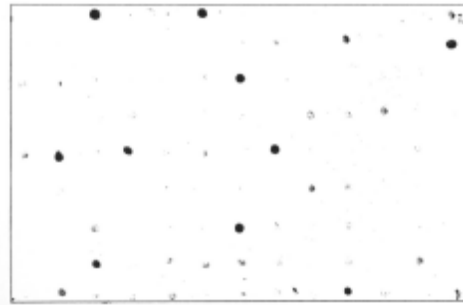
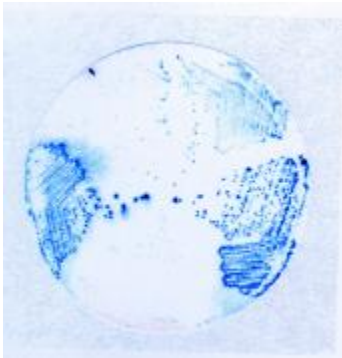
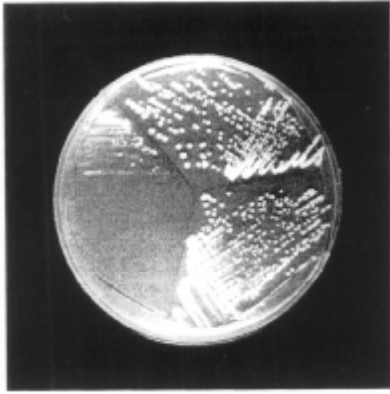


Fig. 3 Electrophoresis of dH731-pBHADNA fragments 2. Yeast two-hybrid screening Fig. 4 Growth assay of yeast for histidine synthesis Fig. 5 X-gal assay for β -galactosidase activity



6 Fig. 6
Selection of library plasmids (Leu²⁺) in *E. coli* HB101
Fig. 7 Dot blotting to reveal identical DNA sequences pBHA-dH731 and pGAD10-library were co-transformed into L40 yeast. The cDNA library from 0-3hr, 3-12hr embryos and larva were used. Through three times of growth and X-gal assays, 133 positive clones which could produce histidine and β -galactosidase were isolated from 5×10^5 successfully transformed yeast clones. These candidates were subdivided into 26 different groups

by dot blotting. Finally, 9 interacting proteins of dH731 were found.
Table 1 Data of the yeast two-hybrid screening

Steps of the yeast two-hybrid screening	Total number of clones
Successful transformants	5,033,600
Interacting clones	245



First test	196
Second test	176
Third test	133

(Each test includes growth (Fig. 4) and X-gal assay (Fig. 5).)



Selection of library plasmids (<i>Leu2⁺</i>) in <i>E. coli</i> HB101 (Fig. 6)	117
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Dot blotting for clone grouping (Fig. 7)	26
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Retransformation for selection true positive interactors	18
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DNA sequencing	9
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dH731 interacting protein	Putative domains & Homologues	Open reading frame (a.a. residue)	Chromosomal localization	Proposed relationship with dH731
1 dH731	Bipartite nuclear localization signal (NLS) (a.a. 57-74)	509	12C	Forming homodimer
2 dUBC9	Ubiquitin-conjugating enzymes active site	158	21C6-7	Assisting dH731 in entering nuclei
3 Ribosomal protein L23a	Bipartite NLS (a.a. 75-92, 96-113), Ala-, Lys-, Pro-rich	269	62B5-10	DNA binding
4 Initiation factor 4a	RNA helicase Vasa	403	67A2-B1	?
5 Novel #103	C ₂ H ₂ zinc finger domain (a.a. 287-318)	321	92B-92C	DNA binding
6 Novel #148	C ₂ H ₂ zinc finger domain (a.a. 144-166)	210	24C3-D2	DNA binding
7 Novel #143	Rat pyrophosphate decarboxylase (DNA sequence identity: 99%)	252	13E-13F	?
8 Novel #415	?	152	66A11-12	?
9 Novel #111	?	?	42D1-E2	?

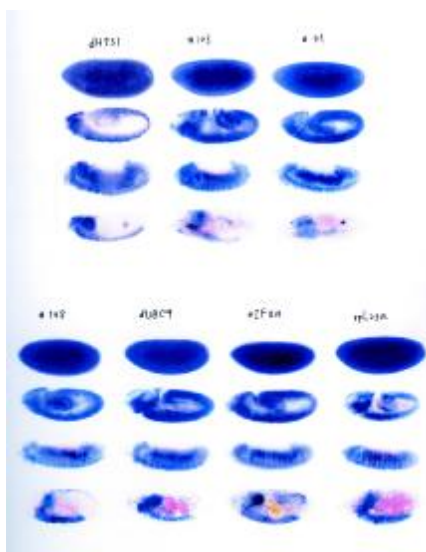


Fig. 8

tains a Bipartite NLS. It is considered that dH731 is able to enter the nucleus because of its NLS, and that dUBC9 is involved in this nuclear import process. In the nucleus, dH731 can interact with rpL23a and two novel proteins, #103 and #148, containing C₂H₂ zinc finger domain isolated in this screening (Table 2). The mRNA of dH731 is highly expressed in embryonic neuroblasts, gonads and the central nervous

4. Discussion By the way of the yeast two-hybrid screening, nine interacting proteins of dH731 were isolated. According to the expression patterns, cellular localization and functions of the dH731-interacting proteins, the possible functions of dH731 were discussed. dUBC9, one of the dH731 interacting proteins, interacts with several transcription factors, such as Tramtrack and Groucho, and is involved in nuclear import. dUBC9 promotes Dorsal, a transcription factor, entering nucleus (5). In dUBC9 mutant, Bicoid, a segment-related transcription factor fails to enter the nucleus and the segment of *Drosophila* embryos is abnormal (6). Moreover, dH731 con

system (CNS) (Fig. 8). These cells are dividing and forming organs in these stages of expression. The mRNA expression patterns of the dH731-interacting genes are all localized in CNS, and partially overlapped in other tissues, such as gonads. Taken together, it is conceivable that dH731 may enter nuclei with the assistance of dUBC9. Within the nucleus, dH731 interacts with the three DNA-binding proteins, rpL23a and two Zn finger proteins (Fig. 10). These interactions may be important for dH731 to play a role in regulation of transcription or cell cycle progression during the neural development and the reproductive organ formation. Fig.10 The model for dH731 entering the nucleus

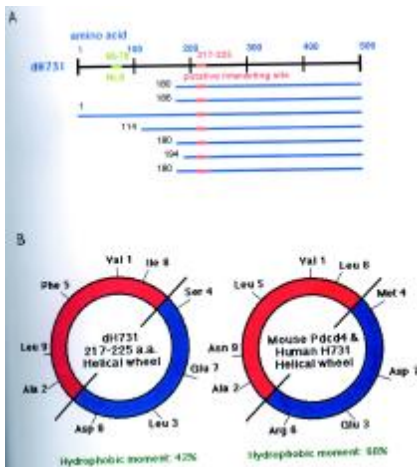


Fig. 9

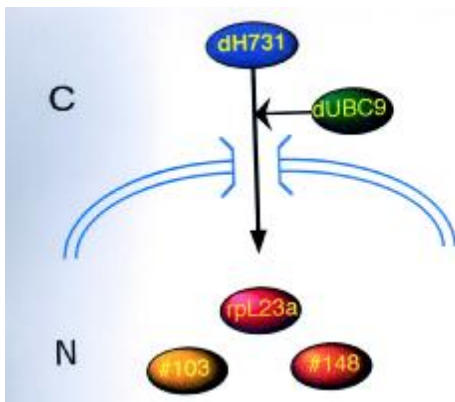


Fig.10

5. References 1. Cmarik J, Min H, Hegamyer G, Zhan S, Kulesz- Martin M, Yashinaga H, Matsubashi S and Colburn N. (1999) Differentially expressed protein Pcd4 inhibits tumor promoter-induced neoplastic transformation. *Proc. Natl Acad. Sci. USA*, 96, 14037-14042. 2. Fields S and Song O-K. (1989) A novel genetic system to detect protein-protein interactions. *Nature*, 340, 245-246. 3. Bhaskar V, Valentine SA and Courey AJ. (2000) A functional interaction between Dorsal and components of the Smt3 conjugating machinery. *J. Biol. Chem.* 275, 4033-4040.