

Sex Organ Development And Interbreeding Mechanism of *Cyathea Spinulosa* (A Tree Fern)

Abstract The study is to investigate the outbreeding mechanisms of *Cyathea spinulosa*, a common Taiwan tree fern (Fig. 1). Fern gametophytes are called prothallia that are known to be bisexual with coexisting archegonia and antheridia. From outdoor surveys, I found that matured prothallia of *C. spinulosa* can be either bisexual or unisexual (contain either archegonia or antheridia). The data also showed that the proportion of female prothallia is much higher in the "isolated" samples (prothallium that is at least 25 mm apart from the closest by neighbor) compared to that of the "clustered" prothallia. In the laboratory, all of the prothallia grown individually could only produce archegonia; however, the prothallia in multispore cultures could be bisexual or unisexual. Induction of antheridium development was studied by placing immature prothallia next to a mature female prothallium. And the induction was both time- and distance-dependent. It takes approximately 8 days for the appearance of the antheridia and the optimal distance is 150-200 mm. Furthermore, prolonged multispore cultures resulted in aborted prothallia (no zygote formation) or sporophyte-carrying prothallia that were originally female or bisexual. These results indicate that: (1) archegonium development is spontaneous, (2) antheridium development must be induced by matured archegonia (from another prothallium), and (3) no antheridium can be induced in archegonia-carrying prothallia; therefore the development of antheridium must precede that of archegonium in an individual prothallium. The above temporal and spatial regulation of sex organ development in *C. spinulosa* gametophytes may represent a mechanism involved in the assurance of outbreeding in this fern.

1. Introduction

Cyathea spinulosa is a common tree fern found in the forests in South east Asia. *C. spinulosa* is believed to be a homosporous fern and thus its gametophytes are believed to be bisexual, i.e., they are hermaphrodites and contain both archegonia and antheridia on a single prothallium (Pal, 1994). Based on the proximity of antheridia and archegonia on bisexual gametophytes, it was once thought that intragametophytic selfing would be the predominant mating system in homosporous ferns (Klekowski, 1979). In a preliminary outdoor survey, I was amazed to find some unisexual prothallia (females or males only) of *C. spinulosa*. The proportion of female prothallia is highly different in samples grown under different growing patterns. It is very high in

isolated samples but very low in clustered samples. After 2 weeks of further culturing the unisexual prothallia in the laboratory, the male prothallia started to develop female organs (archegonia) but the female prothallia remained unchanged. Reviewing of the literature revealed that the mating system of *C. spinulosa* has not been published. I studied the sex ratios of prothallia collected in outdoor surveys and in laboratory cultures with regard to their growing patterns (in "clustered" or "isolated" forms), the time- and distance-dependent induction of antheridia in immature prothallia by older female gametophytes, and the sequence of gametangia formation in *C. spinulosa*. Based on the findings, I conclude that the asynchronous maturation of male and female gametangia can prevent intragametophytic selfing and that the distance-dependent induction of antheridia development may help to promote intergametophytic crossing in this tree fern.

2. Materials and Methods

(Determination of sex expression of *C. Spinulosa* gametophytes in natural habitats)

Cyathea Spinulosa, a common tree fern in Taiwan, was the subject of this study. In three outdoor surveys, prothallia from 2 to 4 mm in radius were collected in two categories according to their growing densities at the Kun-lun Herbal Drug Touring Park in Lungtan, Taoyuan County, Taiwan. The samples were respectively designated as the "clustered prothallia" when they

grew as a group with the farthest individuals less than 2.5 cm away from their neighbors, and the "isolated prothallia" when the closest neighbor was at least 2.5 cm apart. The sex of the samples was determined under a microscope according to the expression of sex organs as shown in Fig. 2. They were further classified into four categories, asexual (no sex organ), male (contained only antheridia), female (contained only archegonia), and hermaphrodite (contained antheridia and archegonia). (Determination of sex expression of *C. Spinulosa* gametophytes in cultures) Segments of leaves bearing matured sporangia collected and stored dried from plants at the Kun-lun Herbal Drug Touring Park in September 1999 were dried and stored in the refrigerator. Spores were then collected and kept at 4°C until used. To establish aseptical gametophyte cultures, spores were surface sterilized with 2.5% NaOCl, rinsed in sterilized water, and sown on 1% solidified agar medium with Bold's solution (Bold, 1957) in 9-cm Petri dishes. The spores were allowed to germinate and grow in a growth chamber, under a 12 hr/day light cycle (fluorescent light, approximately 2,500 lux) at 26 ± 1°C. To simulate the germination condition in the nature habitat, the "clustered gametophytes" cultures were initiated from multispore cultures with the density controlled between 2 to 10 prothallia/cm. In addition, young prothallia (less than 2 mm in radius, 7 to 14 days after germination) were replanted and allowed to grow individually in separated containers to simulate the "isolated gametophytes". For twenty-five days after the spores were sown, the sex of the prothallia was determined as described above. For the determination of sex ratios of sporophyte-carrying gametophytes, multispore cultures were initiated as described above. The cultures were allowed to pro

ceed through the developments of sex organs (maturation), fertilization, and germination of sporophytes. Seventy-five days after the spores were sown, the sporophyte-carrying gametophytes were collected and examined for the presence of still existing or degenerated archegonia and antheridia. The samples were recorded as females and hermaphrodites accordingly. Induction of antheridia in young prothallia Induction of antheridium development was studied by using immature, i.e., asexual prothallia as the recipients and asexual, male as well as female prothallia as the inducers. The prothallia were replanted in containers lined with 4 layers of gauze soaked in Bold's solution. In 9-cm Petri dishes, the inducers were placed at the center and 7 asexual prothallia were placed around the edge (i.e., 4.5 cm apart from the inducer) (Fig. 3A). Development of sex organs was monitored every day after replanting. In

Drug Touring Park, I collected a total of 202 prothallia that grew in clustered configurations (from 7 different locations) and 8 prothallia that grew solitarily. The sex of these prothallia was determined in the laboratory by viewing under a microscope. In addition to an expected great number of hermaphrodites, significant percentages of unisexual prothallia were also found (Fig. 4). Moreover, the ratio of female prothallia in the isolated samples was much higher than that in the clustered samples (42% vs. 5%, as shown in the second column of Figs. 4A and 4B). Subsequent laboratory culture of the unisexual prothallia collected from the wild produced two very different results: the female prothallia remained female but the male prothallia became bisexual. Sex

order to study the distance-dependence of antheridia induction, inducers and recipients were replanted in a 60 x 8 cm glass trough in the following manner: An inducer was placed 5 cm from the edge (starting point) and 4 immature prothallia were placed every 5 cm apart from the inducer (Fig. 3B). Antheridia development in the recipient prothallia was determined 4 days later.

3. Results Sex expression of *C. Spinulosa* gametophytes in natural habitats In three independent visits in the late fall (November to December) to the Kun-lun Herbarial

expression of *C. Spinulosa* gametophytes in cultures To rule out the environmental factors that would affect the sex expression in *C. spinulosa*, spores were allowed to germinate and grow under a controlled environment. In general, the spores germinated within 7 days and took 7 to 14 days to develop into young prothallia, approximately 2 mm in radius and remained asexual. To determine the sex ratios of prothallia grown in different configurations, the sex of the well-developed prothallia was determined 45 days after sowing. Of the 141 prothallia grown in the multispore cultures, 45 (31.9%) of them remained asexual (immature). Moreover, the mature prothallia could be classified as hermaphrodites, female, and male and their ratios were 27.0, 5.7, and 35.5% respectively (Fig. 5A). Therefore, the sex ratio of laboratory-grown prothallia resembles that found in the wild (compared Fig. 4A and Fig. 5A). In contrast, the laboratory-grown solitary pro

thallia were found to be either asexual or female (no hermaphrodite or male). At around 45 days after sowing, the asexual and female prothallia were 71% and 29% respectively (Fig. 5B). Induction of antheridia in young prothallia Although female, male, and asexual prothallia were used respectively as inducers in the induction experiments, only females were found to be effective (Fig. 6). When the distance of the recipient immature prothallia was kept at 4.5 cm as shown in Fig. 3A, the development of antheridia in the recipients took place in a time-dependent manner. Antheridia could be found in some of the recipients as early as 3 days later and all of them developed antheridia 8 days after they were placed in a Petri dish with a female prothallium at the center (Fig. 6A). The induction process was also found to be distance-dependent, the recipients at 15-20 cm away from the inducer were the first to develop antheridia and 100% induction was observed in these samples 4 days after replanting (Fig. 6B). The data showed that 15-20 cm is optimal for the induction process. Sex ratios of sporophyte-carrying gametophytes in prolonged multispore cultures It is conceivable that sporophyte-carrying gametophytes must arise from female or bisexual prothallia. The sex ratio of the sporophyte-carrying gametophytes was determined in the extended multispore cultures. It was found that the majority of the sporophyte-carrying gametophytes were originally hermaphrodites (93%). In other words, only 7% of the sporophyte-carrying gametophytes originated from female prothallia (Fig. 7).

4. Discussion Female prothallia have not yet been found to develop into hermaphrodites in all circumstances while all male prothallia would become hermaphrodites eventually. Therefore, archegonia development in *C. spinulosa* gametophytes is a spontaneous process. In contrast, the development of antheridia in

young prothallia is precisely regulated. The process must be induced by nearby female prothallia that contain mature archegonia. Furthermore, the timing of this induction is critical. It must be initiated before the autonomous development of archegonia in the same prothallium. The developmental sequences of sexual expression of *C. spinulosa* gametophytes are summarized in Fig. 8. As it has been shown in other ferns, mature female *C. spinulosa* gametophytes may release chemical compounds that are collectively called antheridiogens, which act as the chemical inducer for antheridia development on nearby immature prothallia. For each prothallium, whether or not to develop antheridia depends on whether it receives the correct signal(s) at the right time with the right amount. In either case, development of archegonia in these prothallia would take place spontaneously. Moreover, in hermaphrodites, the maturation of archegonia occurred after the sperms were released from the antheridia of the same gametophyte. Finally, a successful fertilization (as indicated by the germination of the sporophytes) may or may not occur (Fig. 8). It is worth noting that only one sporophyte may germinate from one prothallium, indicating that sporophyte development on a gametophyte might be exclusive, it would suppress the development of other zygotes on the same plant.

Although the actions of antheridiogens have been reported extensively, the concentration effect(s) (reflected as the distance-dependent experiment in this study) was not well documented. Under the specified conditions, prothallia at 15-20 cm away from a female gametophyte responded best, development of antheridia in these plants occurred earliest. In nature, spores from the same sporangium are likely to be distributed very closely; therefore it is of interest not to stimulate the closest nearby gametophyte to decrease the chance of intergametophytic selfing.

5. Conclusion In *C. spinulosa*, intragametophytic selfing is most likely prevented by asynchronous maturation of male and female gametangia on the same prothallium and intergametophytic selfing may be prevented by the dose-dependent response of antheridia development to the "not-so-close" female neighbor. Collectively, these mechanisms may evolve to promote intergametophytic crossing (interbreeding) in this tree fern.

6. Reference

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Fig. 1. The morphology of *C. spinulosa* sporophytes and parts. (A) mature *C. spinulosa*; (B) feather-like leaf with leaflets; (C)(D) the ventral side of a leaflet, showing rows of sori; (E) sori; (F) sori; (G) spores.

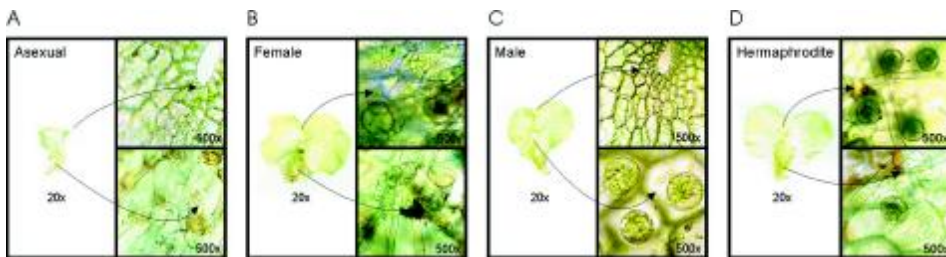
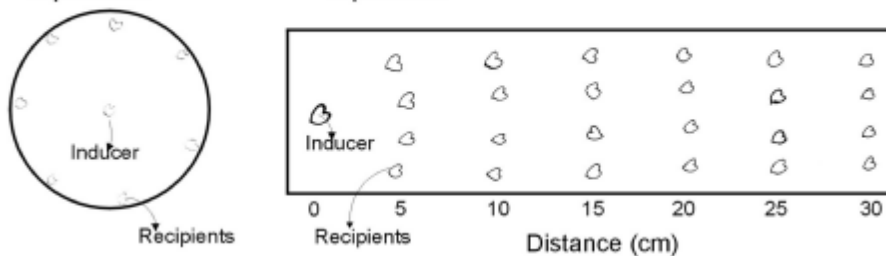


Fig. 2. Sex

expression of *C. spinulosa* gametophytes. Gametophytes of *C. spinulosa* were classified by sexual expressions: (A) asexual gametophyte, no sex organ was found; (B) male gametophyte, prothallium with antheridia; (C) female gametophyte, prothallia were used as recipients.

(A) Time-dependent experiment

(B) Distance-dependent experiment



(A) Time-dependent

experiments, the recipient prothallia were placed 4.5 cm apart from the potential inducers (in 9-cm Petri dishes, the inducers were planted at the center while the recipients were planted at the edge) and the development of antheridia on the recipient prothallia was recorded on a daily basis. (B) Distance-dependent experiments, recipient prothallia were placed every 5 cm apart from the potential inducers and the development of antheridia on the recipient prothallia was recorded on a daily basis.

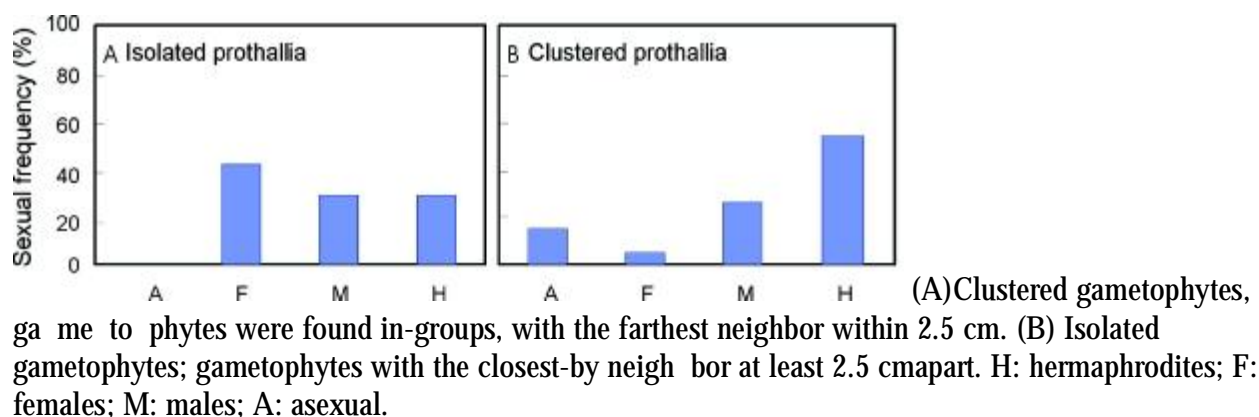


Fig.5. The sexual frequency (%) of *C. spinulosa* gametophytes in the laboratory. (A) Clustered gametophytes, gametophytes that were initiated from multispore cultures, 2-10 gametophytes/cm². (B) Isolated gametophytes, gametophytes that were re-planted after germination and allowed to grow individually, 1 plant per container. H: hermaphrodites; F: females; M: males; A: asexual.

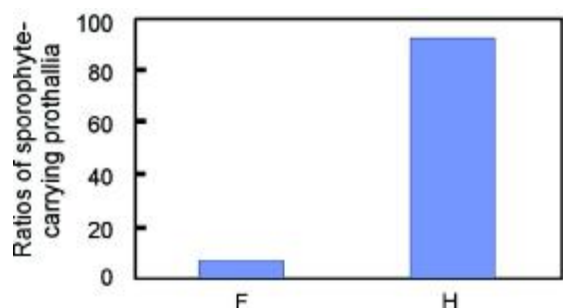


Fig.7 Ratio of female and bisexual

gametophytes with developing sporophytes of *C. spinulosa*. Multispore cultures were initiated and the gametophytes were allowed to grow and develop for 75 days. At the end of this culturing period, young sporophyte-carrying prothallia were collected and observed with an aid of a microscope for the existence of the sex organs. Prothallia containing archegonia (or degenerated archegonia) were classified as female. Prothallia with both archegonia and antheridia (or degenerated organs) were classified as hermaphrodites. H: hermaphrodites; F: females.

